

ZIZYNUMMIN, A DAMMARANE SAPONIN FROM *ZIZYPHUS NUMMULARIA*

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Key Word Index—*Zizyphus nummularia*; Rhamnaceae; dammarane saponin; zizynummin.

Abstract—Zizynummin, a new dammarane saponin isolated from dried leaves of *Zizyphus nummularia*, has been assigned the structure β -D-glucopyranosyl-(1 \rightarrow 2)-6-deoxy- α -L-talopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl-(1 \rightarrow 3)-jubilogenin.

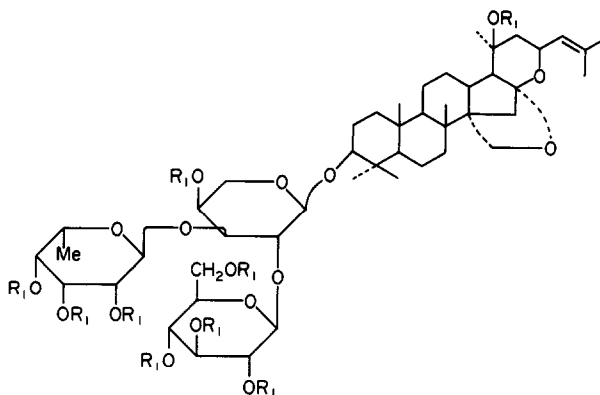
INTRODUCTION

Zizyphus nummularia is a wild growing small shrub which is well-known for its medicinal value [1, 2]. Nummularins A–H and K [3–5], manogenin, taxifolin and taxifolin glycoside [6] have been reported from the plant. A new dammarane saponin is reported here from the ethanolic extract of the leaves.

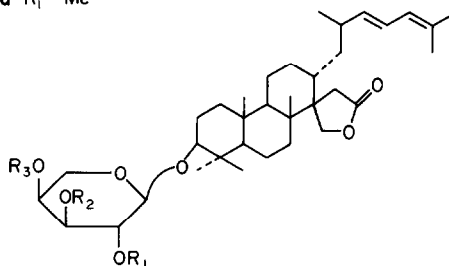
RESULTS AND DISCUSSION

The dried and finely powdered leaves on extraction with ethanol and purification gave a pure saponin which we have named zizynummin (1), mp 255–260°; $[\alpha]_D^{20}$ –44.85°. On acid hydrolysis it yielded ebelin lactone, mp 175–180°, $[\alpha]_D^{20}$ –19.5° (IR, UV and EIMS) and L-arabinose, D-glucose, and 6-deoxy-L-talose (co-PC). However, the IR and UV of Zizynummin showed the absence of a lactone ring and conjugated double bond, respectively, therefore, ebelin lactone must be an artefact

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1 $R_1 = H$
1a $R_1 = Me$



- 2 $R_1 = \beta$ -D-Glucose, $R_2 = R_3 = H$
2a $R_1 = 2,3,4,6$ -Tetra-*O*-methyl β -D-glucose, $R_2 = R_3 = Me$
3 $R_2 = 6$ -Deoxy- α -L-talose, $R_1 = R_3 = H$
3a $R_2 = 2,3,4$ -Tri-*O*-methyl-6-deoxy- α -L-talose, $R_1 = R_3 = Me$

of the real aglycone. Smith-de Mayo degradation [7] (performed twice) of **1** yielded jujubogenin, mp 249–251°, $[\alpha]_D^{20} -31.8^\circ$, (mmp, co-TLC, EIMS and comparison of ^1H NMR) and this compound appears to be the real aglycone of **1**.

The FDMS of **1** showed a cationized cluster ion at m/z 935 $[\text{M} + \text{Na}]^+$ (base peak) and the fragment ion peaks at m/z 789 $[(\text{M} + \text{Na}) - 146]^+$, 773 $[(\text{M} + \text{Na}) - 162]^+$ and 627 $[(\text{M} + \text{Na}) - 308]^+$ suggested that **1** was in a pure state and had a branched chain trisaccharide composed of methyl pentosyl-(hexosyl)-pentose in the ratio of 1:1:1 combined with a compound of MW 471 (jujubogenin-H) [8]. In addition, characteristic ions at m/z 163 (glucose-OH) and at 147 (6-deoxy-L-talose-OH) were also displayed (Fig. 1) which further proved that two different terminal sugars, namely methyl pentose and hexose, were present in **1** [9]. The ^1H NMR signals of three anomeric protons at δ 4.20 (1H, *d*, $J = 5.5$ Hz), 4.26 (1H, *d*, $J = 7.0$ Hz) and 5.28 (1H, *br s*), indicated that the L-arabinose possesses an α -, D-glucose a β - and 6-deoxy-L-talose an α -linkage, which were also supported by the application of Klyne's rule [10].

Methylation of **1** by the modified Hakomori method [11, 12] furnished nona-*O*-methylether, **1a**, which on methanolysis gave a mixture of methyl pyranosides of 2,3,4,6-tetra-*O*-methyl- β -D-glucose, 2,3,4-tri-*O*-methyl-6-deoxy- α -L-talose and 4-mono-*O*-methyl- α -L-arabinose (GC). Hydrolysis of the above sugar mixture gave corresponding methylated sugars (PC). 4-Mono-*O*-methyl- α -L-arabinose was positive to Wallenfels' reagent [13]. The EIMS fragments at m/z 219, 187 $[219 - \text{MeOH}]^+$ and 189, 157 $[189 - \text{MeOH}]^+$ also confirmed the presence of β -D-glucose and 6-deoxy- α -L-talose as the terminal sugars and also that one of them was attached at C-2 and the other at C-3 of α -L-arabinose which in turn was linked at C-3 with jujubogenin.

Partial hydrolysis of zizyminin afforded two propapogenins **2** and **3**. The UV spectrum showed that **2** and **3** possess an open chain conjugated triene system [14]. Hydrolysis of **2** furnished ebelin lactone, L-arabinose and D-glucose but **3** afforded 6-deoxy-L-talose instead of D-glucose (co-PC). Hydrolysis of **2** and **3** permethyl ethers, prepared by a modified Hakomori's method, afforded 2,3,4,6-tetra-*O*-methyl- β -D-glucose with 3,4-di-*O*-methyl- α -L-arabinose (also positive to Wallenfels' reagent) and 2,3,4-tri-*O*-methyl-6-deoxy- α -L-talose along with 2,4-di-*O*-methyl- α -L-arabinose, respectively (PC). Moreover, treatment of **3** with sodium periodate liberated free L-arabinose (co-PC).

Therefore, **2** and **3**, formed from the splitting of 6-deoxy-L-talose and D-glucose, respectively, from **1**, possess the structures β -D-glucopyranosyl-(1 \rightarrow 2)- α -L-arabi-

nopyranosyl-(1 \rightarrow 3)-ebelin lactone and 6-deoxy- α -L-talopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl-(1 \rightarrow 3)-ebelin lactone and, hence, zizyminin can be assigned the structure β -D-glucopyranosyl-(1 \rightarrow 2)-6-deoxy- α -L-talopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl-(1 \rightarrow 3)-jujubogenin.

EXPERIMENTAL

Mps are uncorr. PC was carried out on Whatman No. 1 paper using the descending method and aniline hydrogen phthalate as the visualizing agent. The solvent systems used were: (A) CHCl_3 -MeOH- H_2O (65:25:10); (B) *n*-BuOH-HOAc- H_2O (4:1:5); (C) *n*-BuOH-EtOH- H_2O (5:1:4); (D) C_6H_6 -Me₂CO (10:1); (E) C_6H_6 -EtOAc (4:1). GC of methylated sugars; column-succinate polyester (10% H.P.), N_2 (30 ml/min), programmed at 175° at 6°/min.

Isolation and purification of zizyminin. Dried and finely powdered leaves of *Zizyphus nummularia*, collected from Meerut (U.P.), were successively extracted with petrol (40–60°), EtOAc and finally with 95% EtOH ($\times 3$, 8 hr). The EtOH extract was dried *in vacuo* and partitioned between H_2O and H_2O satd *n*-BuOH (1:1, $\times 3$). The *n*-BuOH layer was treated with 5% aq. K_2CO_3 soln (twice) and then washed with H_2O . The concd *n*-BuOH extract was subjected to repeated CC (solvent A) to give zizyminin (**1**, 3 g; mp 255–260° (MeOH); $[\alpha]_D^{20} -44.85^\circ$ (MeOH; *c* 1.00); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400, 1400; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: no absorption above 200. FDMS (Fig. 1); ^1H NMR (100 MHz, DMSO- d_6): δ 3.09 (1H, *br s*, OH-20) 4.20 (1H, *d*, $J = 5.5$ Hz), 4.26 (1H, *d*, $J = 7.0$ Hz) and 5.28 (1H, *br s*).

Acid hydrolysis of 1. A soln of **1** (100 mg) in MeOH was hydrolysed by refluxing with 10% aq. HCl (15 ml) for 4 hr, cooled, diluted with H_2O and filtered to afford the ebelin lactone (20 mg). Colourless needles, mp 175–180° (MeOH); $[\alpha]_D^{20} -19.5^\circ$; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3500, 1770, 1642, 1600; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 268 (4.58), 278 (4.67) and 288 (4.55); EIMS (probe) 15 eV, m/z (rel. int.): 454 $[\text{M}]^+$ (0.6), 436 (15), 207 (20), 189 (40), 121 (52), 43 (100). The neutralized (Ag_2CO_3) and concd aq. hydrolysate showed the presence of three sugars only, viz. D-glucose, L-arabinose and 6-deoxy-L-talose (co-PC, authentic samples run in parallel, solvent B, R_f 0.18, 0.21 and 0.44, respectively).

Smith-de Mayo degradation. Compound **1** (500 mg) was oxidized with NaIO_4 (1 g in 100 ml at 50% aq. EtOH) at room temp. for 46–60 hr in the dark with continuous stirring. The reaction mixture was refluxed with 5% KOH soln for 4 hr and then extracted with *n*-BuOH ($\times 3$). The final product, obtained after removal of solvent, was submitted to CC (solvent D) to yield pure sapogenin (160 mg). Jujubogenin, colourless needles, mp 249–251° (MeOH); $[\alpha]_D^{20} -31.8^\circ$ (EtOH; *c* 0.8); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400 (OH, *br*), 1400; EIMS (probe) 15 eV, m/z (rel. int.): 472 $[\text{M}]^+$ (0.4), 454 $[\text{M} - \text{H}_2\text{O}]^+$ (19.4), 207 (14.2), 191 (14.7), 189 (16.9), 121 (34.2), 109 (67.1), 97 (100), 43 (41.5); ^1H NMR (200 MHz,

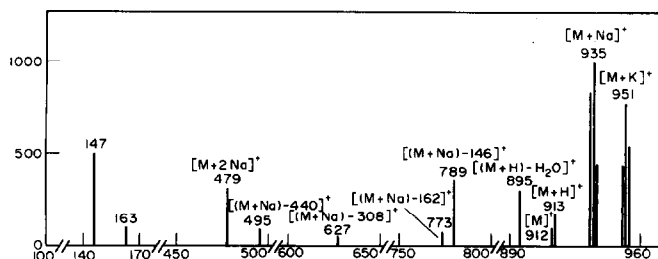


Fig. 1. FDMS of compound **1** between 20 and 45 mA emitter heating current.

C_5D_5N : δ 0.80, 0.95, 1.10, 1.15, 1.33 (3H each, s), 1.68 (6H, br s), 2.38 (1H, d, $J = 8$ Hz), 2.76 (1H, q-like), 3.35 (1H, t, $J = 8$ Hz), 4.14 (2H, $2 \times d$, $J = 15$ Hz), 4.30 (1H, br), 5.35 (1H, br d, $J = 7$ Hz) and 5.70 (1H, br s, OH).

Permethylation of 1. Compound 1 (300 mg) was permethylated by the modified Hakomori method to yield the nona-*O*-methyl derivative 1a, which was purified (CC, solvent E), mp 93.96°; $[\alpha]_D^{20} -48.5^\circ$ (CHCl₃; c 0.6); IR ν_{max}^{KBr} cm⁻¹: no-OH; EIMS (probe) 15 eV, m/z (rel. int.): 833 [$M - \text{tri-}O\text{-methyl-6-deoxy-L-talose} + H$]⁺ (0.3), 803 [$M - \text{tetra-}O\text{-methyl-glucose} + H$]⁺ (0.4), 454 (4.4), 219 (10.9), 189 (100), 187 (92.5), 157 (70.8), 155 (13.7), 101 (100), 88 (100) and 45 (25.6). (Found: C, 64.28; H, 9.10. $C_{56}H_{94}O_{17}$ requires: C, 64.71; H, 9.12%.)

Methanolysis of 1a. Compound 1a (100 mg) in 1 N HCl-MeOH (15 ml) was refluxed (3 hr), neutralized (Ag₂CO₃), diluted, filtered and the concd filtrate was subjected to GC, R_f (min): 2,3,4,6-tetra-*O*-methyl- β -D-glucose (2.02); 2,3,4-tri-*O*-methyl-6-deoxy- α -L-talose (2.87); 4-mono-*O*-methyl- α -L-arabinose (8.59). Hydrolysis of a portion of the above sugar mixture with 10% HCl showed the presence of the following sugars on PC (solvent C); 4-mono-*O*-methyl-L-arabinose, 2,3,4-tri-*O*-methyl-6-deoxy-L-talose and 2,3,4,6-tetra-*O*-methyl-D-glucose (R_G : 0.38, 0.88 and 1.0, respectively). On spraying with Wallenfels' reagent the spot corresponding to R_G value 0.38 gave an intense pink colour.

Partial hydrolysis. A soln of zizunummin (1g) in 5% aq. HCl-MeOH (1:1, 50 ml) was heated under reflux on a steam bath for 45 min. The reaction mixture was neutralized with Ag₂CO₃ and filtered. The filtrate was evaporated to dryness *in vacuo* and the residue was separated by CC (solvent A) to give ebelin lactone (50 mg) and two prosapogenins. Compound 2 (150 mg); mp 260–265° (MeOH); UV λ_{max}^{MeOH} nm (log ϵ): 266 (4.58), 276 (4.67) and 288 (4.55). Compound 3 (175 mg) mp 255–260° (MeOH); IR ν_{max}^{KBr} cm⁻¹: 3400, 1765, 1640, 1600; UV λ_{max}^{MeOH} nm (log ϵ): 267 (4.58), 277 (4.67) and 288 (4.55). Compounds 2 and 3 (25 mg) were separately hydrolysed and worked-up in the usual way to yield ebelin lactone (mmp, co-TLC and IR) and neutral concd aq. hydrolysates of 2 and 3 showed D-glucose (R_f 0.18) and L-arabinose (R_f 0.21); L-arabinose and 6-deoxy-L-talose (R_f 0.44), respectively (co-PC with authentic samples, solvent B).

Permethylation and methanolysis of 2 and 3. Portions (75 mg) of both 2 and 3 were separately permethylated by the modified Hakomori method to give their per-*O*-methyl derivatives 2a and 3a, respectively. Compound 2a: mp 78–80°; IR ν_{max}^{KBr} cm⁻¹: no OH; EIMS (probe) 15 eV, m/z (rel. int.): 832 [M]⁺ (0.4), 597 (0.3), 219 (20), 187 (80), 88 (100). Compound 3a mp 87–90°; IR ν_{max}^{KBr} cm⁻¹: no OH; EIMS (probe) 15 eV, m/z (rel. int.): 802 [M]⁺ (0.3), 597 (0.6), 189 (100) and 157 (22). Compounds 2a and 3a were

methanolysed followed by hydrolysis and work-up in the usual way. The methyl sugars from 2a were identified (PC, solvent C) as 3,4-di-*O*-methyl-L-arabinose (R_G 0.65), positive to Wallenfels' reagent, and 2,3,4,6-tetra-*O*-methyl-D-glucose (R_G 1.0). Compound 3a gave 2,4-di-*O*-methyl-L-arabinose (R_G 0.65) and 2,3,4-tri-*O*-methyl-6-deoxy-L-talose (R_G 0.88).

Periodate oxidation of 3. Compound 3 (25 mg) in H₂O (10 ml) was mixed with NaIO₄ (250 mg) and the soln kept in the dark for 48 hr. Ethylene glycol (1 ml) was added to decompose excess NaIO₄ and the soln was hydrolysed with 10% MeOH-HCl (45 min). Its filtrate was neutralized, concd and examined by PC (solvent B) to detect L-arabinose (R_f 0.21) only.

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